

Learning Objectives

- Identify different assays for minimal residual disease (MRD) detection
- List diseases in which MRD monitoring is considered the standard of care
- Define potential uses of MRD monitoring

Topics

- Minimal Residual Disease
 - Definition
 - Assays
 - Relevance
- MRD in CML
- MRD in Myeloma
- MRD in AML
- MRD in ALL
- Summary

Lets go back in time...

- The year is 1995
- 32-year-old female
- Has recently been experiencing increasing fatigue and weight loss
- WBC 48,000 with 10% myeloblasts in peripheral blood
- and 5% basophils

 Bone marrow 10% blasts; 5% basophils
- Cytogenetic studies show t(9;22) with an additional 20q- abnormality
- Has a 39-year-old sister who is a 6/6 HLA match

Case Presentation

- 3 months post allograft she is in a complete cytogenetic remission.
- 18 months later a new test called PCR is reported as positive
- · She continues to be followed.
- At 24 months the tests is negative.
- At 30 months post BMT the test is positive again
- What should be done (remember it is 1998 now)

CML: ARS Question #1

In patients with CML a QUALITATIVE PCR (pos or neg) predicts relapse.

- 1. True
- 2. False

CML: ARS Question #2

In this patient what would the appropriate next step be with this positive test for minimal residual disease (MRD)?

1. Request donor lymphocyte infusion (DLI) as soon as possible

2. Start interferon

3. Wait until 1999 and get her on a protocol with the new STI571

4. Perform bone marrow aspiration and determine whether there was cytogenetic evidence of disease

ARS Question #3: What is MRD anyway?

Which is the correct definition for MRD?

 Minimal residual disease refers to disease that is left over after treatment that only can be seen by an expert pathologist.
 Minimal residual disease only relates to CML and represents presence of disease at a 1 in 100000 level.

3.Minimal residual disease is the name given to small numbers of leukemic or other tumor cells detected by very sensitive methods that remain in the patient during treatment, or after treatment when the patient is in remission. It is the major cause of relapse in cancer and leukemia.

MRD-Definition When in doubt ask WIKIPEDIA

 "Minimal residual disease is the name given to small numbers of leukaemic cells that remain in the patient during treatment, or after treatment when the patient is in remission. It is the major cause of relapse in cancer and leukaemia."

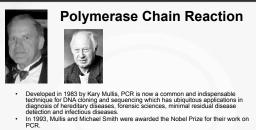
Minimal Residual Disease

- · Not totally true
- MRD has usually referred to disease detected by non-traditional methods (xray or pathology). The two most commonly used methods are flow cytometry and polymerase chain reaction.
- MRD by flow cytometry or PCR predicts for a higher risk of relapse after chemotherapy and also after transplantation in SOME but NOT ALL diseases.

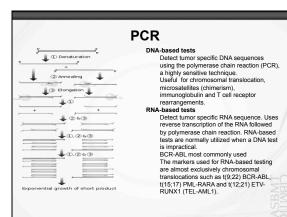
MRD Detection

- Cytogenetic methods, including FISH

 Generally not sensitive enough to be real <u>minimal</u> residual disease measure
- Flow cytometry
 - Based on aberrant antigen expression ("Leukemiaassociated immunophenotype")
- PCR
 - Adaptable to different targets
 - Can measure clonal abnormality or abnormal expression



- PCR. The method relies on thermal cycling, consisting of cycles of repeated heating and cooling of the reaction for DNA metling and enzymatic replication of the DNA. Primers (short DNA fragments) containing sequences complementary to the target region along with a DNA Polymerase
 - Almost all PCR applications employ a heat-stable DNA polymerase, such as Taq polymerase (an enzyme originally isolated from the bacterium Thermus aquaticus



Types of PCR Methods

- Antigen receptor PCR
 Most suited to lymphoid malignancies
- Fusion transcript PCR
 Several tumor types but only limited subsets of most tumors (CML excepted)
- PCR for gene mutations
 AML subsets, e.g. FLT3 or NPM1
- mRNA PCR
 - Suitable for upregulated genes, e.g. WT1

Immunological tests

 Flow cytometry is an immunological-based testing of leukemias or other cancers utilizes proteins on the surface of the cells. Leukemic and other cancer cells often show quite unusual and unique combinations (leukemic phenotype) of these cell surface proteins. These proteins can be stained with fluorescent dye labeled antibodies and detected using <u>flow cytometry</u>. The limit of detection of immunological tests is generally about 1 in 10,000 cells and cannot be used on cancers that don't have an identifiable and stable phenotype

Flow Cytometry-WIKIPEDIA

- Flow cytometry is a laser-based, biophysical technology employed in cell counting, sorting, biomarker detection and protein engineering.
- Principle: suspend cells in a stream of fluid and passing them by an electronic detection apparatus. It allows simultaneous multi-parametric analysis of the physical and chemical characteristics of up to thousands of particles per second.
- Flow cytometry is routinely used in the diagnosis of health disorders, especially blood cancers.
- . History
- Mack Fulwyler was the inventor of the forerunner to today's flow cytometers Wolfgang Gohde developed in 1968 fluorescent based flow cytometry

- Patient-specific testing
- Patient-specific MRD detection using immunoglobulin (IG) or T cell receptors (TCR). •
- Immunoglobulin (IG) or 1 cell receptors (TCR). Measures MRD in tumors that do not contain a chromosomal translocation or other specific marker. These tests are very specific, and detect leukaemic cells at levels down to one cell in a million, though the limit typically achieved is 1 in 10,000 to 1 in 100,000 cells. For comparison, the limit of what one can detect using traditional morphologic examinations using a microscope is about 1 cell in 100. 100.

Methods For Detecting Chimerism

XY FISH

- Easy, but not very sensitive; only applicable to a subset of patients
- PCR methods
 - Microsatellite markers(short tandem repeats (STRs) or variable number tandem repeats (VNTR))
 Informative in nearly all cases; sensitivity around 1-5%
 - · Most widely used
 - TaqMan qPCR against single nucleotide polymorphisms · Sensitivity 0.1% or better, and quantitation better but perhaps not informative as often; more limited data
 - Y chromosome PCR even more sensitive (1/10⁵)
- · Lineage-specific chimerism more specific

General Considerations

Two broad approaches

- Chimerism
 - · Not a direct measure of disease · Applicable to all patients
 - · Extent of chimerism not necessarily related to disease burden
- Minimal residual disease detection
 - · Genetic approaches to detect actual clone (PCR, FISH) Phenotypic approaches to detect abnormal expression (flow cytometry, mRNA)
- Imaging and other clinical monitoring discussed in manuscript

CML-Audience Response Questions

False

- Patients with CML may have low level QUALITATIVE PCR without ever relapsing
- · In this patient what would the appropriate next step be with this positive test for minimal residual disease (MRD)?
- · Perform bone marrow aspiration and determine whether there was cytogenetic evidence of disease.

Comparison of Flow Cytometry and PCR for MRD Detection

PCR

FLOW Advantages Rapid and relatively

inexpensive, allowing early intervention

- Widely applicable in many

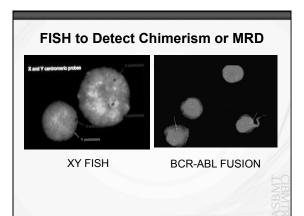
diseases (not CML)

- Advantages Highly sensitive and reproducible
 Clone specific
 - Most data in many diseases
- Disadvantages

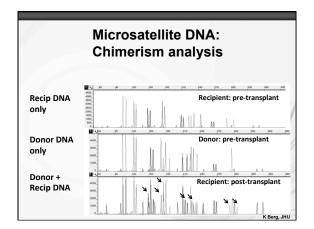
 Not applicable to all diseases

- useases Ag receptor PCR requires allele specific oligos and is expensive and time consuming Clonal evolution a potential pitfall
- Not well standardized

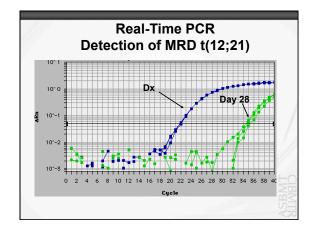
 Disadvantages - Not as sensitive as PCR







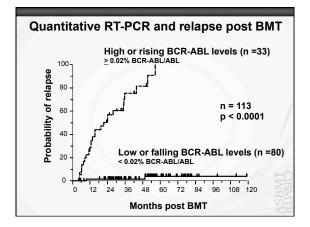




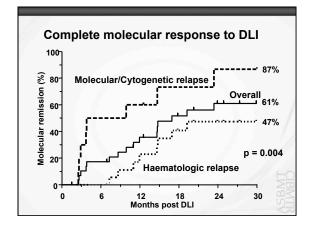


| Quar 10 ¹² - 10 ¹⁰ - 10 ¹⁰ - 10 ⁸ - 10 ⁶ - 10 ⁶ - 10 ⁴ - 10 ⁶ - | CHR Cytogenetic CCR Cytogenetic response CCR 3 log reduction Q-PCR 4 log reduction Complete Molecular | CML 100 10 BCRABL/ABL 0.01 BL ratio (%) 0.0001 (%) |
|---|--|--|
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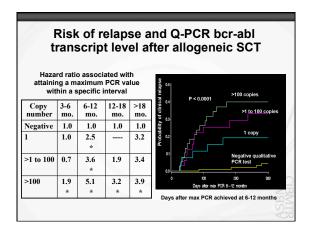




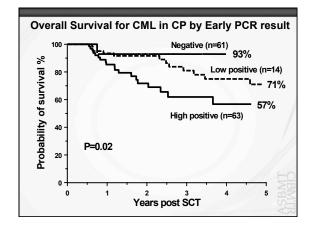


| | | Hem.Rel→Death | Non-Hem.Rel → Death |
|--------------------|--------------|-------------------|---------------------|
| EDMT sale | Low | 1.00 | 1.00 |
| EBMT risk score | Intermediate | 1.42 (0.62-3.26) | 1.14 (0.58-2.21) |
| SCOLE | High | 2.99 (0.92-5.72) | 4.35 (1.86-10.20) |
| Occurrence | 1993-1996 | 7.42 (3.07-17.97) | 3.50 (1.79-6.81) |
| of relapse | 1997-1999 | 8.14 (3.50-18.94) | 2.34 (1.15-4.76) |
| in | 2000-2003 | 6.54 (2.46-17.37) | 0.60 (0.22-1.64) |





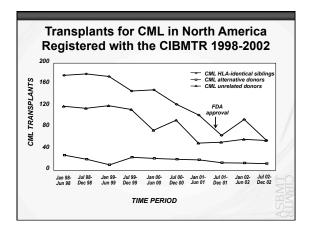






CML Summary

- MRD monitoring is well established with Q-PCR for BCR-ABL
- PCR positivity predicts for relapse (all types) and disease progression
- PCR monitoring can be used for assessing response to treatment of relapse (DLI +/- Imatinib)
- Treatment of early relapse (molecular-cytogenetic)
 results in superior response rates and survival
- There is a need for standardization of PCR methodology
- Future clinical trials should focus on MRD monitoring after treatment with TKIs post allogeneic SCT

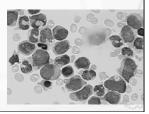


| DISEASE | PCR | FLOW |
|------------------|---------------------|--------------------|
| ALL | Yes | Yes |
| AML | Subgroups only | Yes |
| CML | Yes | No |
| CLL | Yes | Yes, probably best |
| NHL | Yes | Limited data |
| Myeloma* | Yes | Yes |
| Hodgkin lymphoma | No | No |
| MPN | Limited data (JAK2) | No |



NCI SCT Relapse Monitoring Subcommittee -Monitoring strategies in Acute Myeloid Leukemia and Myelodysplastic Syndromes

Ulrike Bacher, MD Clinic for Stem Cell Transplantation University Cancer Center Hamburg Germany

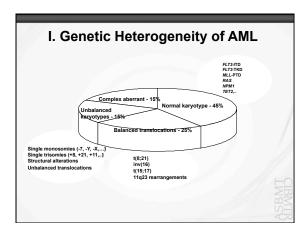


Case Presentation

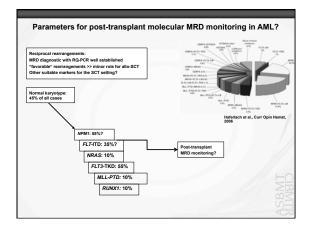
- · 55 year old male with relapsed acute leukemia
- Has an t(8;21) and initial remission lasted 24
 months
- Undergoes an allogeneic SCT.
- 18 months post SCT in in a hematologic and cytogenetic remission but PCR is still reported as positive
- What should be done (remember it is 2014 now)

ARS #4: What to do?

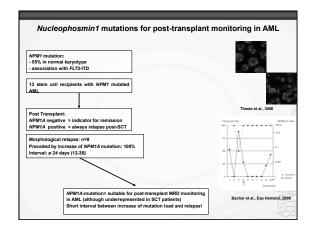
- 1. Proceed to 2nd allo SCT
- 2. Proceed to DLI
- 3. Proceed to chemo with cytarabine
- 4. Continue to monitor with PCR and treat only if increasing levels of disease or evidence of hematologic relapse develops



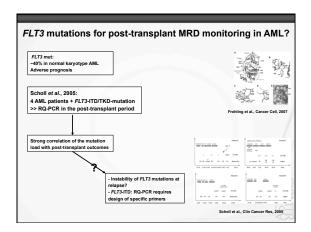




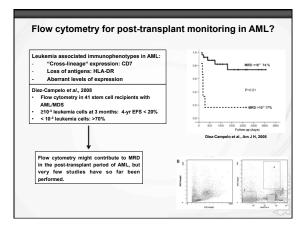




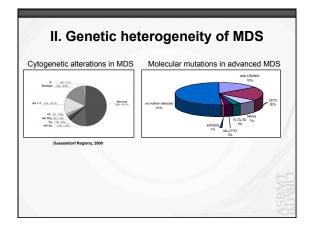




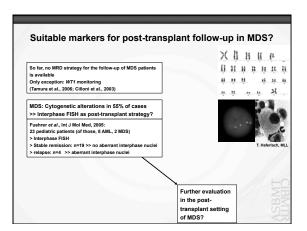




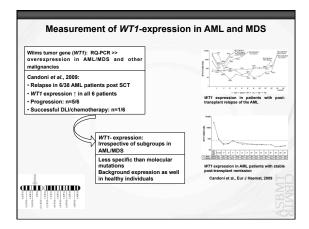




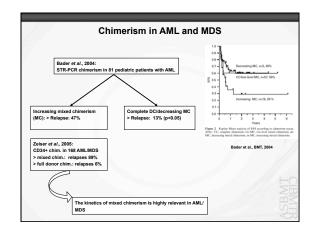












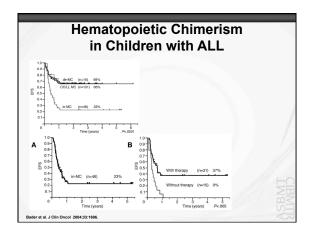


Disease-Specific Methods and Strategies for Monitoring Relapse Following Allogeneic Stem Cell Transplantation

Pediatric Acute Lymphoblastic Leukemia

on behalf of the Sub-Committee

Peter Bader, Wendy Stock, Andre Willasch, Alan Wayne



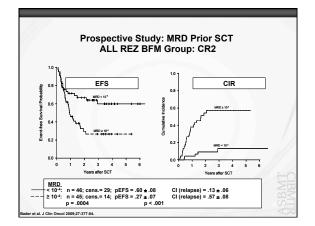


| | | dies o and Int | | | m |
|------------------------------------|--------------------------|--------------------------------|--|---------|--|
| Author | Number of patients | Diagnosis | Interval of investigations | Methods | Relapses |
| Formakova Haematologica 2003 | 54 | AL, CML and MDS children | weekly to +100; monthly | STR | MC associated with rejection and relapse Immunotherapy was possible |
| Gorczynska BMT 2004 | 14 | ALL, AML children | weekly to +100; monthly | STR | In-MC could be converted by immunotherapy to CC |
| Bader JCO 2004 | 163 | ALL children | weekly to +100; monthly | STR | MC associated with rejection and relapse Immunotherapy was possible |
| Horn BMT 2008 | 20 | AL children | 1,3,6,12 months; In MC bi- weekly | STR | MC associated with relapse IT was not possible |

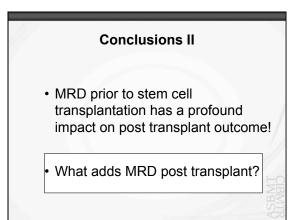


| Retros | pectiv | | dies - M erature | IRD pr | ior to SCT |
|--------------------------------------|--------------------|-----------|--------------------------|-----------------|--|
| Author | Number of patients | Diagnosis | Time of investigation | Methods | Survival according to MRD status |
| Knechtli Blood 1998 | 64 | ALL | prior to conditioning | Ig / TCR PCR | high level pos. – 0% low level pos. – 36% negative – 73% |
| Bader Leukemia 2002 | 41 | ALL | prior to conditioning | Ig / TCR PCR | high level pos. – 23% low level pos. – 48% negative – 78% |
| Uzunel Blood 2001 | 30 | ALL | prior to conditioning | Ig / TCR PCR | high level pos. – 47% low level pos. – 50% negative – 100% |
| Sramkova Ped Blood Cancer 2007 | 25 | ALL | prior to conditioning | Ig / TCR PCR | positive – 0% negative – 94% |







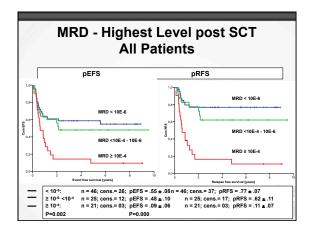


| Retros | spect | | tudies - terature | | Post SCT |
|----------------------|--------------------------|-----------|-------------------------------------|-------------------|--|
| Author | Number of patients | Diagnosis | Time of investigation | Methods | Survival and MRD status |
| Knechtli BJH 1998 | 68 | ALL | up to 24 months post SCT | Ig / TCR PCR | relapse – 88% pos. remission – 22% pos. |
| Uzuel BJH 2003 | 23 | ALL | 24 months | Ig / TCR PCR | MRD pos. associated with relapse |
| Sanchez BJH 2002 | 40 | ALL | d30, 60, 90, every 2-3 months | Flow cytometry | positive - 33% negative - 74% |



| | spective S BFM Grou | • | |
|-------------------|------------------------|--------------|----------|
| N | | 92 | |
| Diagnosis | | ALL | |
| Remission | | ≥ CR2 | |
| Transplant Period | Jan 19 | 99 | May 2006 |
| Evaluation | Ja | nuary 15th 2 | 2009 |
| Follow up | Median | Min | Max |
| [Years] | 5.13 | 3.44 | 6.48 |







Conclusions III and Summary

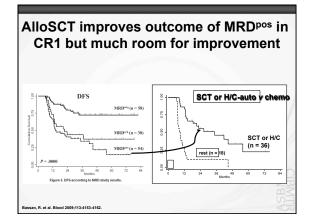
- MRD assessment in BM post transplant is predictive for relapse
 - Serial BM investigations are warranted.
 - Current working recommendations of the BFM: days 30, 60, 100, 200, 300, 365, at 18 months and 24 months.

<u>Summary:</u>

- Patients with mixed chimerism have a high risk for relapse
- Patients, who become/remain MRD positive $\geq 10^{-4}$, have a very high risk to develop relapse · Additional treatment in these patients is warranted

MRD in adults with ALL

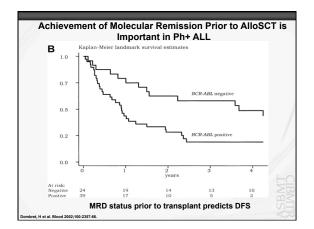
- Shown to be useful predictor of DFS in many studies (non-transplant) •
 - Independent prognostic feature - Mostly using PCR techniques - IgH/TCR, fusion genes
 - "Informative" assay available in 60-90% of patients
- Early CR time-points predictive of outcome: from 4-22 weeks following initiation of treatment
- Fewer studies evaluating role of MRD in setting of alloSCT



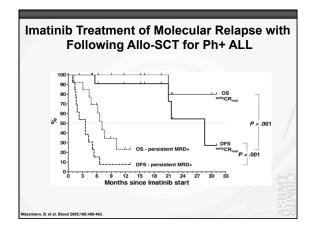


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|--|--------------------------|--|-----------------------|---|--|
| Author | Number of patients | Diagnosis | Time of investigation | Methods | DFS and MRD status |
| Mortuza JCO 2002 | 19 | ALL (B-lineage) | From 1-20 mos. | Ig / TCR PCR Semi-quant. | positive – 0% negative – 100% CCR |
| Spinelli Haematologica 2007 | 37 | ALL | Day +100 | Ig/TCR or fusion gene PCR Quantitative | positive >10 ^{-4:} 20% negative: 93% |
| Bassan* Blood 2009 | 18 | ALL *All were PCR+ prior to transplant | Not defined | Ig / TCR PCR | positive >10 ⁻⁴ : 0 negative: 50% |







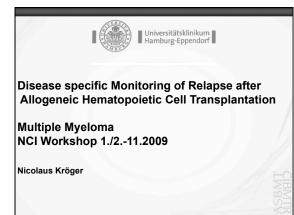




Summary

- MRD detection both prior to and following alloSCT for adults with ALL is associated with poor DFS •
- Clinical interventions based on MRD measurements suggest utility but data are very limited: Allocation to alloSCT in CR1 •

 - Post-transplant intervention to prevent relapse
 Targeted therapy (e.g. imatinib) following transplant
- Challenge: implementation of standardized MRD assays that can be done in "real-time" ٠
 - IgH/TCR qPCR assays are laborious Data on flow cytometric measurements of MRD in adults with ALL are lacking



Conventional techniques for monitoring

- · Bone marrow aspiration: infiltration often underestimated
- Serum/24h urine electrophoresis (agarose gel or capillary zone): lowest detectable level of M-component: 0.2 - 0.6 g/L
- Immunofixation (serum/urine): lowest detectable level of M-component: 0.12 - 0.25 g/L
- Free light chain assay (κ/λ ratio) : useful in light chain disease and non-secretory, necessary to determine sCR, early response assessment due to short half time (6h)

Imaging monitoring

- More than 80% of the pts develop osteolytic bone lesions
- The hallmark of myeloma bone disease is an increased osteoclastic bone resorption and an exhausted osteoblast function resulting in a reduced bone formation even in patients in complete remission
- Standard: conventional radiology as skeletal survey involving cervical, thoracic and lumbar spine, skull, chest, pelvis, humeri and femora
- Disadvantage: low sensitivity, no exact response assessment
- CT: high sensitivity, but higher radiation dose
- MRI: high sensitivity, no radiation dose, detect
 extramedullary disease
- PET-CT: highest sensitivity for extramedullary disease

Flow-cytometry

- Flow cytometry has become an easy applicable method to detect residual myeloma cells The European Myeloma Network recommends a minimal panel including
- CD19, CD56, CD20, CD117, CD28 and CD27.
- Plasma cell gating should be based on CD38 vs. CD138 expression
- This method is less sensitive (10⁻⁴) than allelespecific oligonucleotides PCR (ASO-PCR)

A.C., et al. Haematologica 2008;93:431-43

Allele-specific oligonucleotides PCR (ASO-PCR)

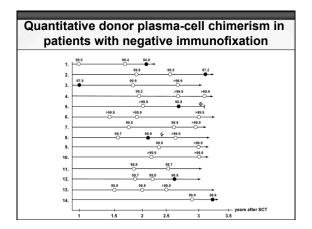
- Patient-specific primers (IgH rearrangement)
- High sensitivity of (10⁻⁵ 10⁻⁶) and highly specific (100%)
- Time-consuming (for each patients), does not detect extramedullary disease

| on re | arranged immu chain ge | noglobulin heavy enes |
|--------|---------------------------|--------------------------------------|
| In CR: | • | 50% molecular CR 7% molecular CR |
| | | |
| In CR: | | 50% molecular CR 16% molecular CR |

| tem cen tran | splantatio | n | |
|----------------|------------|----------------|---------|
| ıltiple Myelo | ma (EBMT-S | Studie): Pat w | vith CR |
| | PCR neg | PCR mixed | PCR pos |
| No. of pts | 16 | 19 | 13 |
| 5 year cumulat | tiv | | |
| sk of relapse | 0% | 33% | 100 |

Chimerisms

- Not specific for relapse, in majority of relapse donor cell chimerism persisted
- Lineage specific chimerism (plasma cell-chimerism: CD138+ BM cells)
- By using real-time PCR the sensitivity of the method is 10⁻⁴ to 10⁻⁵. The disadvantage of the methods is the lack of specificity.



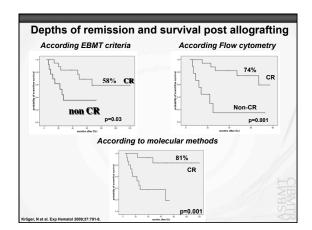


Predictive value of donor-plasma-cell chimerism for relapse

- 93% with stable or increasing donor-plasmacell chimerism remained immunofixationnegative.
- 83% with a decrease of donor-plasma-cell chimerism was associated with relapse in the sense of becoming immunofixation-positivity (in 2: 3 and 6 months prior than immunofixation becomes positive)

tol 2006:34:688-94.

N et al. Exp Her





Summary

- MRD assessment is now routinely performed in the setting of hematologic malignancies.
- MRD presence can predict disease recurrence in some but not all instances.
- Although frequently done the impact of early intervention based on MRD assessment has only been shown to be effective in CML.
- Both patients and physicians should be encouraged to participate in clinical trials.